## CHAPTER 13

# Health Benefits of n-3 Polyunsaturated Fatty Acids: Eicosapentaenoic Acid and Docosahexaenoic Acid

Nalin Siriwardhana,\*\*<sup>†,1</sup> Nishan S. Kalupahana,\*\*<sup>†,‡</sup> and Naima Moustaid-Moussa\*\*<sup>†</sup>

Contents	I.	Sources and Intakes of Eicosapentaenoic Acid and	
		Docosahexaenoic Acid	212
	II.	Health Benefits of n-3 PUFAs	213
		A. n-3 PUFAs and cardiovascular diseases	214
	III.	Anti-inflammatory Effects of EPA and DHA	215
	IV.	n-3 PUFAs and Metabolic Disorders	216
	V.	Health Concerns	218
	References		219

#### Abstract

Marine-based fish and fish oil are the most popular and well-known sources of n-3 polyunsaturated fatty acids (PUFAs), namely, eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA). These n-3 PUFAs are known to have variety of health benefits against cardiovascular diseases (CVDs) including well-established hypotriglyceridemic and anti-inflammatory effects. Also, various studies indicate promising antihypertensive, anticancer, antioxidant, antidepression, antiaging, and antiarthritis effects. Moreover, recent studies also indicate anti-inflammatory and insulin-sensitizing effects of these

<sup>\*</sup> Department of Animal Science, University of Tennessee, Knoxville, Tennessee, USA

<sup>†</sup> UT Obesity Research Center, University of Tennessee, Knoxville, Tennessee, USA

Department of Physiology, Faculty of Medicine, University of Peradeniya, Peradeniya, Sri Lanka

<sup>&</sup>lt;sup>1</sup> Corresponding author: Nalin Siriwardhana, E-mail address: rsiriwar@utk.edu

fatty acids in metabolic disorders. Classically, n-3 PUFAs mediate some of these effects by antagonizing n-6 PUFA (arachidonic acid)induced proinflammatory prostaglandin E2 (PGE2) formation. Another well-known mechanism by which n-3 PUFAs impart their anti-inflammatory effects is via reduction of nuclear factor-κΒ activation. This transcription factor is a potent inducer of proinflammatory cytokine production, including interleukin 6 and tumor necrosis factor-α, both of which are decreased by EPA and DHA. Other evidence also demonstrates that n-3 PUFAs repress lipogenesis and increase resolvins and protectin generation, ultimately leading to reduced inflammation. Finally, beneficial effects of EPA and DHA in insulin resistance include their ability to increase secretion of adiponectin, an anti-inflammatory adipokine. In summary, n-3 PUFAs have multiple health benefits mediated at least in part by their anti-inflammatory actions; thus their consumption, especially from dietary sources, should be encouraged.

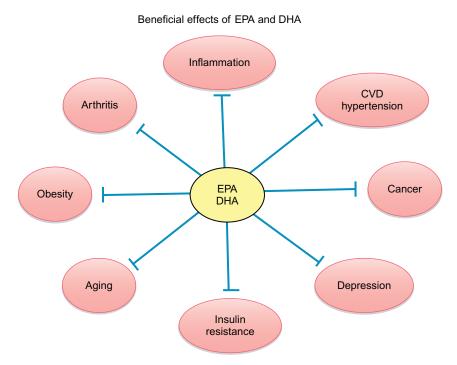
### I. SOURCES AND INTAKES OF EICOSAPENTAENOIC ACID AND DOCOSAHEXAENOIC ACID

Marine fish is the well-known source of eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA). EPA (C<sub>20</sub>H<sub>30</sub>O<sub>2</sub>; 20:5n-3) and DHA  $(C_{22}H_{34}O_2; 22:6n-3)$  are long-chain  $\omega$ -3/n-3 polyunsaturated fatty acids (PUFAs). Technically, EPA is a nonessential n-3 fatty acid as the human body can convert essential n-3 alpha-linolenic acid (ALA) into EPA and DHA. However, in humans, this conversion is not efficient enough to meet the EPA and DHA demand to impart beneficial health effects; thus, it is expected to obtain these fatty acids from dietary sources. The n-3 PUFA levels consumed in the United States and many other countries in the world are very low compared to the recommended intake of n-6/n-3 PUFAs (11:1 (n6:n3) vs. 2.3:1; Davis and Kris-Etherton, 2003; Whelan and Rust, 2006). EPA and DHA are among the most studied bioactive compounds of marine origin. Fish oil being the major and well-known source of EPA and DHA, dietary fish such as anchovy, bluefish, herring, mackerel, mullet, sardines, salmon, sturgeon, tuna, and trout are popular sources of fish oil. Further, due to its recognized health benefits and recommendation by several health agencies, fish oil (primarily DHA and EPA) has become a very popular dietary supplement. Fish oil is commercially available as soft gel capsules. These preparations are often available with added vitamins, antioxidants, and various flavors. EPA and DHA are not only found in fish oil but also in marine algal species. However, they are considerably less concentrated in most of the algae, compared to some fish sources. Most of the identified n-3 PUFA-rich algae contain higher amount of DHA compared to EPA.

Fat is a key component in the human diet. Research shows that excessive consumption of saturated fat negatively impacts several biomarkers of health while monounsaturated and n-3 PUFAs are beneficial to human health. Moreover, research shows that unbalanced dietary ratios of n-6: n-3 may lead to various health complications as well as disease progression while increased n-3 levels impart prevention and health promoting effects (Burghardt *et al.*, 2010; Goodstine *et al.*, 2003; Simopoulos, 2002; Wan *et al.*, 2010). The American Heart Association recommends eating fatty fish meals at least twice a week due to their promising health and especially cardiovascular benefits. Here, we review some of the health benefits of n-3 PUFAs, due in part to their anti-inflammatory effects in cancer, cardiovascular diseases (CVDs), obesity, and other metabolic disorders.

#### **II. HEALTH BENEFITS OF N-3 PUFAS**

Fish oil-derived n-3 PUFAs have been a healthy nutritional supplement for long time due to numerous health benefits including promising effects to improve cardiovascular health (Harris, 2007; Lu et al., 2011), brain function, and overall health during pregnancy (Koletzko et al., 2008). Also, fish oil-derived n-3 PUFAs are well known to prevent inflammation (Calder, 2008; Fetterman and Zdanowicz, 2009; Figueras et al., 2011), aging (Dyall et al., 2010), arthritis (Wall et al., 2010), insulin resistance (Kalupahana et al., 2010a), and depression and slows the progression of certain cancers (Astorg, 2004; Cabanes et al., 2003; Leitzmann et al., 2004; Fig. 13.1). Further, it has been shown that fish oil-derived n-3 PUFAs can prevent cardiac remodeling and dysfunction under pressure overload conditions in rats (Duda et al., 2009) and atrial fibrillation associated with heart failure in a rabbit (Kitamura et al., 2011). Also, EPA and DHA are known to induce breast cancer cell apoptosis and impart anticancer effects. Wu et al. (2005) showed that in vivo and in vitro treatment of n-3 PUFAs can inhibit the breast cancer growth through activation of a neutral sphingomyelinase-mediated pathway (Wu et al., 2005). Moreover, it has been reported that an approach with fish oil nutritional intervention can improve the health in lung cancer patients (Murphy et al., 2011). In a recent study, we reported that EPA significantly prevents and reverses insulin resistance in high-fat diet-induced obese mice via modulation of adipose tissue inflammation (Kalupahana et al., 2010a). Moreover, it has been reported that supplementation of long-chain ω-3 PUFAs in elderly female patients reduces the occurrence of depressive symptoms, improves phospholipid fatty acid profile and health-related quality of life (Rondanelli et al., 2011).



**FIGURE 13.1** Beneficial effects of EPA and DHA. Scientifically validated beneficial effects of EPA and DHA as evident by epidemiological evidence as well as *in vivo* and *in vitro* studies. Blocked lines are to indicate the suppression effects. EPA, eicosapentaenoic acid; DHA, docosahexaenoic acid; CVD, cardiovascular disease.

n-3 PUFAs are mostly known for their consistent hypotriglyceridemic effects (Harris, 1996, 1997). Moreover, diets rich in PUFAs are able to alter adipose tissue gene expression and metabolism. Hypotriglyceridemic effects of n-3 PUFA are believed to be primarily mediated via peroxisome proliferator activated receptor (PPAR $\alpha$ )-dependent mechanisms (Buettner *et al.*, 2006), which may also account for the improved insulin resistance and reduction of hepatic steatosis in people consuming diets high in n-3 PUFA.

#### A. n-3 PUFAs and cardiovascular diseases

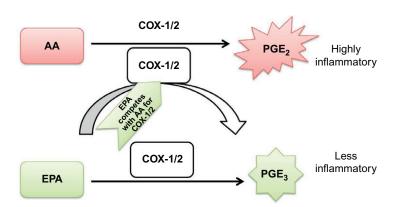
n-3 PUFAs reduce the risk of sudden cardiac death. For example, in the Physicians Health Study, individuals with the highest n-3 PUFA level in blood had a 90% reduction in sudden cardiac death compared to ones with the lowest n-3 PUFA levels (Albert *et al.*, 2002). These fatty acids also

reduce the total mortality and sudden death in patients following myocardial infarction (Marchioli *et al.*, 2002). These beneficial effects of n-3 PUFA in reducing sudden cardiac death are attributed to two main factors. First, n-3 PUFAs reduce cardiac arrhythmias (Raitt *et al.*, 2005), which are responsible for the majority of sudden cardiac deaths. Second, these fatty acids increase atherosclerotic plaque stability (Thies *et al.*, 2003), thereby preventing plaque rupture and subsequent coronary events.

n-3 PUFAs also reduce several risk factors for CVDs. These include reductions in plasma triglyceride levels as well as several inflammatory cytokine levels. Detailed anti-inflammatory mechanisms of n-3 PUFA are discussed below.

#### III. ANTI-INFLAMMATORY EFFECTS OF EPA AND DHA

One of the main anti-inflammatory mechanisms is that EPA acts as a competitive inhibitor for proinflammatory arachidonic acid (AA) on cyclooxygenase (COX), which produces proinflammatory eicosanoids (Fig. 13.2). When EPA is subjected to oxygenation by COX, the less-inflammatory prostaglandin  $E_3$  (PGE<sub>3</sub>) is generated while AA generates highly proinflammatory prostaglandin  $E_2$  (PGE<sub>2</sub>). As EPA is structurally similar to AA, EPA can also compete with AA to incorporate into the cell membrane phospholipids and become the preferential substrate for COX



**FIGURE 13.2** EPA competitively inhibits PGE<sub>2</sub> formation by COX-1 and COX-2. EPA competitively binds with COX-1 and COX-2 and produces less-inflammatory PGE<sub>3</sub> while suppressing AA binding which produces highly inflammatory PGE<sub>2</sub>. AA, arachidonic acid; COX, cyclooxygenase; EPA, eicosapentaenoic acid; PGE<sub>2/3</sub>, prostaglandin E<sub>2/3</sub>. Calder (2009) was used as reference for the design.

activity. Oxygenation of AA by COX can be inhibited by n-3 PUFAs and PGE<sub>3</sub> is generated instead of PGE<sub>2</sub>. Thus, n-3 PUFAs act like nonsteroidal anti-inflammatory drugs to decrease COX enzyme activities and subsequently decrease inflammation including in adipose tissue as indicated by our recent studies (Ferrucci et al., 2006; Kalupahana et al., 2010a; Wortman et al., 2009). Indeed, treatment of EPA in the diet can decrease PGE<sub>2</sub> levels through competitive inhibition of AA (Lands et al., 1973). Also, it has been shown that EPA and DHA can suppress the COX-2 activity in human umbilical vein endothelial cells (Lee et al., 2009). Thus, PGE<sub>2</sub> levels can be manipulated by increasing consumption of EPA. COX, also known as prostaglandin H<sub>2</sub> synthase (PGH<sub>2</sub> synthase), has both COX and peroxidase activity. The COX complex is made up of COX-1 and COX-2 enzymes. COX-1 is constitutively expressed while the expression of COX-2 is inducible and highly regulated. Pharmacological inhibition of COX-2 with celecoxib decreases PGE<sub>2</sub> secretion in adipocytes, and subsequently, lipolysis can be also decreased (Wortman et al., 2009). Therefore, when COX-associated biochemical reactions are intensively associated with EPA due to increased EPA levels and decreased proinflammatory AA levels, less harmful and significantly reduced inflammatory status can be expected. Thus, increased n-3 PUFA levels compared to n-6 PUFA can impart significant beneficial health effects against inflammation. Also, it has been recently suggested that increased n-3 PUFA levels in patients can improve the outcome of critically ill patients (Ott et al., 2011).

#### IV. N-3 PUFAS AND METABOLIC DISORDERS

In both animals and humans, n-3 PUFA was shown to reduce inflammation and insulin resistance (Bierhaus et al., 2004; Jones et al., 2003, 2005). Both EPA and DHA are known to have anti-inflammatory activities that reduce the proinflammatory mediators such as tumor necrosis factor-α (TNF-α), interleukin 1 (IL-1), IL-6, IL-8, resistin, plasminogen activator inhibitor 1, and monocyte chemotactic protein-1 (MCP-1) levels (Kalupahana et al., 2010b). Obesity is associated with a chronic low-grade inflammation characterized by an increase in proinflammatory cytokines and adipokines. EPA and DHA are well known to beneficially alter several metabolic processes including carbohydrate and lipid metabolism. n-3 PUFAs increase β-oxidation of fatty acids and inhibit hepatic and adipose lipogenic gene transcription (Al-Hasani and Joost, 2005; Jones et al., 1996; Sampath and Ntambi, 2004), thus shifting lipids from synthetic pathways to oxidative pathways. Further, dietary n-3 PUFAs act as ligand activators of PPARa, which induces the transcription of genes involved in β-oxidation (Neschen et al., 2002). Lipidmetabolizing genes downregulated by PUFAs include the fatty acid synthase, lipoprotein lipase, hormone-sensitive lipase, adipocyte lipid-binding protein (aP2), acetyl-CoA carboxylase, and the enzyme stearoyl-CoA desaturase-1 (Jones *et al.*, 1996; Sessler *et al.*, 1996).

Also, it was recently demonstrated that saturated fatty acids can activate Toll-like receptor 4 (TLR4) and TLR2 signaling in adipocytes leading to increased transcription of inflammatory markers via with nuclear factor-κB (NF-κB). NF-κB activation has been implicated in many inflammatory processes (Lira et al., 2010). Some proinflammatory cytokines such as TNF-α and IL1 directly activate NF-κB. NF-κB activation has been reported to play a pivotal role in obesity, cancer, atherosclerosis, diabetes, arthritis, and other CVD, inflammatory, and immunological disorders (Bierhaus et al., 2004; Jones et al., 2003, 2005). It has been suggested that NF-κB may be an important therapeutic target for specific CVDs (Bierhaus et al., 2004; Jones et al., 2005; Ruan and Lodish, 2003). MCP-1, which plays a critical role in the development of CVD, is increased by activated NF-κB (Niu and Kolattukudy, 2009; Zhu et al., 2008). Moreover, MCP-1 can induce adipogenesis, macrophage infiltration into adipose tissue, and insulin resistance (Kanda et al., 2006; Younce et al., 2009; Zhu et al., 2008). Several studies have reported that NF-kB activation can be suppressed by fish oil and also identified that suppression of NF-κB activation has been strongly associated with mechanisms responsible for suppression of several disease conditions by fish oil (Encarnacion et al., 2008, 2011; Lee et al., 2009; Suzuki et al., 2010; Zhao et al., 2004). Further, Bellenger et al. (2011) showed that pancreatic n-3 fatty acid enrichment inhibits proinflammatory cytokines and NF-κB protein expression and increases IκBα protein expression in MLD-STZ-induced fat-1 transgenic mice.

It has been shown that fish oil can not only suppress proinflammatory mediators but also can increase the anti-inflammatory ones such as adiponectin (Duda *et al.*, 2009; Kalupahana *et al.*, 2010b). Increased adiponectin levels can reduce inflammation and beneficially improve the metabolism. Specifically, the increased levels of adiponectin can significantly reduce the insulin resistance. Oster *et al.* (2010) showed that DHA increases cellular adiponectin mRNA and secreted adiponectin protein in 3T3-L1 adipocytes, possibly by a mechanism involving PPAR $\gamma$ . A recent dietary intervention study conducted on healthy Japanese female subjects by Kondo *et al.* (2010) showed that a fish-based diet intervention increased the serum adiponectin concentration in young, nonobese, healthy Japanese female subjects. Also, the same study indicated that the increment in serum  $\omega$ -3 PUFA may regulate the serum adiponectin concentration (Kondo *et al.*, 2010).

Other reported mechanisms mediating PUFA effects in metabolic disorders include binding to GPCR-120 and generation of newly discovered resolvins and protectins (Ariel and Serhan, 2007; Schwab *et al.*, 2007). Stimulation of GPR120 with n-3 FAs causes broad anti-inflammatory

effects in monocytic RAW 264.7 cells and in primary intraperitoneal macrophages (Oh *et al.*, 2010). Further, it has been reported that inefficient biosynthesis of n-3-derived mediators of resolution of inflammation in muscle and adipose tissue contributes to the maintenance of chronic inflammation in obesity (White *et al.*, 2010). Hellmann *et al.* (2011) showed that resolving D1 can decrease adipose tissue macrophage accumulation and improves insulin sensitivity in obese-diabetic mice.

#### V. HEALTH CONCERNS

It is always a concern that heat, light, and oxygen can promote fish oil degradation which can produce harmful compounds such as peroxides. With the advances in food and pharmaceutical science, antioxidants are now incorporated into fish oil to prevent fish oil oxidation. Also, the effective storage and packaging methods have been developed to assure the quality. However, it is important to follow the storage guides provided with the products and simply avoid the exposure to heat, light, and oxygen. Also, extended cooking and deep frying can make fish oil unhealthy. Moreover, the quality and source of the fish oil is highly important. Some fish and fish oil may have unhealthy levels of heavy metals such as mercury and selenium. Some studies have reported that fish oil contaminated with mercury can increase CVD risk instead of expected CVD protection benefits while some studies indicate that there should be intensive investigations to confirm the harmful effects of mercury on CVD (Mozaffarian et al., 2011; Virtanen et al., 2005). Specially, when eating fish or taking fish oil supplements during pregnancy, it is always good to carefully select fish oil source for the safety of both the mother and the baby.

Taken together, n-3 PUFAs such as EPA and DHA are promising health promoting compounds and intake of dietary EPA and DHA can suppress the inflammation and inflammation-associated health complications. While there are several mechanisms suggested and detailed for the anti-inflammatory activities of EPA and DHA, here we have summarized a few important mechanisms such as competitive inhibition of AAmediated PGE<sub>2</sub> formation, suppression of NF-κB activation, generation/ production/secretion of anti-inflammatory mediators (adiponectin, resolving, and protectin), and anti-inflammatory activities mediated via GRP120. Also, EPA and DHA are known to beneficially alter the metabolic processes by reducing adiposity and increasing lipid oxidation, thus contributing antiobesity effects in general. Moreover, the reduction of excessive adiposity can directly reduce the low-grade chronic inflammation associated with obesity. Therefore, EPA and DHA can also be used in dietary interventions effectively to overcome obesity, inflammation, and many other disease conditions.

#### REFERENCES

- Albert, C. M., Campos, H., Stampfer, M. J., Ridker, P. M., Manson, J. E., Willett, W. C., and Ma, J. (2002). Blood levels of long-chain n-3 fatty acids and the risk of sudden death. *N. Engl. J. Med.* **346**, 1113–1118.
- Al-Hasani, H. and Joost, H. G. (2005). Nutrition-/diet-induced changes in gene expression in white adipose tissue. *Best Pract. Res. Clin. Endocrinol. Metab.* **19**, 589–603.
- Ariel, A. and Serhan, C. N. (2007). Resolvins and protectins in the termination program of acute inflammation. *Trends Immunol.* **28**, 176–183.
- Astorg, P. (2004). Dietary n-6 and n-3 polyunsaturated fatty acids and prostate cancer risk: A review of epidemiological and experimental evidence. Cancer Causes Control 15, 367–386.
- Bellenger, J., Bellenger, S., Bataillem, A., Massey, K. A., Nicolaou, A., Rialland, M., Tessier, C., Kang, J. X., and Narce, M. (2011). High pancreatic n-3 fatty acids prevent STZ-induced diabetes in fat-1 mice: Inflammatory pathway inhibition. *Diabetes* **60**, 1090–1099.
- Bierhaus, A., Humpert, P. M., and Nawroth, P. P. (2004). NF-kappa B as a molecular link between psychosocial stress and organ dysfunction. *Pediatr. Nephrol.* **19**, 1189–1191.
- Buettner, R., Parhofer, K. G., Woenckhaus, M., Wrede, C. E., Kunz-Schughart, L. A., Schölmerich, J., and Bollheimer, L. C. (2006). Defining high-fat-diet rat models: Metabolic and molecular effects of different fat types. J. Mol. Endocrinol. 36, 485–501.
- Burghardt, P. R., Kemmerer, E. S., Buck, B. J., Osetek, A. J., Yan, C., Koch, L. G., Britton, S. L., and Evans, S. J. (2010). Dietary n-3:n-6 fatty acid ratios differentially influence hormonal signature in a rodent model of metabolic syndrome relative to healthy controls. *Nutr. Metab. (Lond)* 7, 53.
- Cabanes, A., Wang, M., Olivo, S., Gustafsson, J., and Hilakivi-Clarke, L. (2003). Effect of n-3 polyunsaturated fatty acids (PUFAs) on breast cancer progression. *Cancer Epidemiol. Biomarkers Prev.* **12** (1305S).
- Calder, P. (2008). The relationship between the fatty acid composition of immune cells and their function. *Prostaglandins Leukot. Essent. Fatty Acids* **79**, 101–108.
- Calder, P. C. (2009). Polyunsaturated fatty acids and inflammatory processes: New twists in an old tale. *Biochimie*. **91**, 791–795.
- Davis, B. and Kris-Etherton, P. (2003). Achieving optimal essential fatty acid status in vegetarians: Current knowledge and practical implications. *Am. J. Clin. Nutr.* **78**, 640S–646S.
- Duda, M., O'Shea, K., Tintinu, A., Xu, W., Khairallah, R., Barrows, B., Chess, D., Azimzadeh, A., and Harris, W. (2009). Fish oil, but not flaxseed oil, decreases inflammation and prevents pressure overload-induced cardiac dysfunction. *Cardiovasc. Res.* **81**, 319–327.
- Dyall, S., Michael, G., and Michael-Titus, A. (2010). Omega-3 fatty acids reverse age-related decreases in nuclear receptors and increase neurogenesis in old rats. *J. Neurosci. Res.* **88**, 2091–2102.
- Encarnacion, M., Warner, G., Gray, C., Cheng, J., Keryakos, H., Nath, K., and Grande, J. (2008). Signaling pathways modulated by fish oil in salt-sensitive hypertension. Am. J. Physiol. Renal Physiol. 294, F1323–F1335.
- Encarnacion, M., Warner, G., Cheng, J., Gray, C., Nath, K., and Grande, J. (2011). n-3 Fatty acids block TNF-alpha-stimulated MCP-1 expression in rat mesangial cells. *Am. J. Physiol. Renal Physiol.* 300, F1142–F1151.
- Ferrucci, L., Cherubini, A., Bandinelli, S., Bartali, B., Corsi, A., Lauretani, F., Martin, A., Andres-Lacueva, C., Senin, U., and Guralnik, J. M. (2006). Relationship of plasma polyunsaturated fatty acids to circulating inflammatory markers. J. Clin. Endocrinol. Metab. 91, 439–446.
- Fetterman, J. and Zdanowicz, M. (2009). Therapeutic potential of n-3 polyunsaturated fatty acids in disease. *Am. J. Health Syst. Pharm.* **66**, 1169–1179.

- Figueras, M., Olivan, M., Busquets, S., Lopez-Soriano, F., and Argiles, J. (2011). Effects of eicosapentaenoic acid (EPA) treatment on insulin sensitivity in an animal model of diabetes: Improvement of the inflammatory status. *Obesity* 19, 362–369.
- Goodstine, S. L., Zheng, T., Holford, T. R., Ward, B. A., Carter, D., Owens, P. H., and Mayne, S. T. (2003). Dietary (n-3)/(n-6) fatty acid ratio: Possible relationship to premenopausal but not postmenopausal breast cancer risk in U.S. women. *J. Nutr.* **133**, 1409–1414.
- Harris, W. S. (1996). n-3 fatty acids and lipoproteins: Comparison of results from human and animal studies. *Lipids* **31**, 243–252.
- Harris, W. S. (1997). n-3 fatty acids and serum lipoproteins: Animal studies. *Am. J. Clin. Nutr.* **65**, 1611S–1616S.
- Harris, W. S. (2007). Omega-3 fatty acids and cardiovascular disease: A case for omega-3 index as a new risk factor. *Pharmacol. Res.* **55**, 217–223.
- Hellmann, J., Tang, Y., Kosuri, M., Bhatnagar, A., and Spitem, M. (2011). Resolvin D1 decreases adipose tissue macrophage accumulation and improves insulin sensitivity in obese-diabetic mice. FASEB J. 25, 2399–2407.
- Jones, B. H., Maher, M. A., Banz, W. J., Zemel, M. B., Whelan, J., Smith, P. J., and Moustaid, N. (1996). Adipose tissue stearoyl-CoA desaturase mRNA is increased by obesity and decreased by polyunsaturated fatty acids. Am. J. Physiol. 271, E44–E49.
- Jones, W. K., Brown, M., Ren, X., He, S., and McGuinness, M. (2003). NF-kappaB as an integrator of diverse signaling pathways: The heart of myocardial signaling? *Cardiovasc. Toxicol.* 3, 229–254.
- Jones, W. K., Brown, M., Wilhide, M., He, S. W., and Ren, X. P. (2005). NF-kappa B in cardiovascular disease—Diverse and specific effects of a "general" transcription factor? *Cardiovasc. Toxicol.* 5, 183–201.
- Kalupahana, N. S., Claycombe, K., Fletcher, S., Wortman, P., and Moustaid-Moussa, N. (2010a). Eicosapentaenoic acid improves adipose tissue inflammation in part via downregulation of adipose angiotensinogen secretion. *Obesity* 18 (S71-S).
- Kalupahana, N. S., Claycombe, K., Newman, S. J., Stewart, T., Siriwardhana, N., Matthan, N., Lichtenstein, A. H., and Moustaid-Moussa, N. (2010b). Eicosapentaenoic acid prevents and reverses insulin resistance in high-fat diet-induced obese mice via modulation of adipose tissue inflammation. J. Nutr. 140, 1915–1922.
- Kanda, H., Tateya, S., Tamori, Y., Kotani, K., Hiasa, K. I., Kitazawa, R., Kitazawa, S., Miyachi, H., and Maeda, S. (2006). MCP-1 contributes to macrophage infiltration into adipose tissue, insulin resistance, and hepatic steatosis in obesity. J. Clin. Invest. 116, 1494–1505.
- Kitamura, K., Shibata, R., Tsuji, Y., Shimano, M., Inden, Y., and Murohara, T. (2011). Eicosapentaenoic acid prevents atrial fibrillation associated with heart failure in a rabbit model. Am. J. Physiol. Heart Circ. Physiol. 300, H1814–H1821.
- Koletzko, B., Lien, E., Agostoni, C., Böhles, H., Campoy, C., Cetin, I., Decsi, T., Dudenhausen, J. W., and Dupont, C. (2008). The roles of long-chain polyunsaturated fatty acids in pregnancy, lactation and infancy: Review of current knowledge and consensus recommendations. J. Perinat. Med. 36, 5–14.
- Kondo, K., Morino, K., Nishio, Y., Kondo, M., Fuke, T., Ugi, S., Iwakawa, H., Kashiwagi, A., and Maegawa, H. (2010). Effects of a fish-based diet on the serum adiponectin concentration in young, non-obese, healthy Japanese subjects. J. Atheroscler. Thromb. 17, 628–637.
- Lands, W. E., Letellierm, P. R., Romem, L. H., and Vanderhoekm, J. Y. (1973). Inhibition of prostaglandin biosynthesis. Adv. Biosci. 9, 15–28.
- Lee, S., Kim, H., Chang, K., Baek, J., Park, J., Shin, J., Choi, W., Lee, J., and Paik, W. (2009). DHA and EPA down-regulate COX-2 expression through suppression of NF-kappa B activity in LPS-treated human umbilical vein endothelial cells. *Korean J. Physiol. Pharmacol.* 13, 301–307.

- Leitzmann, M., Stampfer, M., Michaud, D., Augustsson, K., Colditz, G., Willett, W., and Giovannucci, E. (2004). Dietary intake of n-3 and n-6 fatty acids and the risk of prostate cancer. *Am. J. Clin. Nutr.* **80**, 204–216.
- Lira, F. S., Rosa, J. C., Pimentel, G. D., Tarini, V. A., Arida, R. M., Faloppa, F., Alves, E. S., do Nascimento, C. O., and Oyama, L. M. (2010). Inflammation and adipose tissue: Effects of progressive load training in rats. *Lipids Health Dis.* 9, 109.
- Lu, J., Borthwick, F., Hassanali, Z., Wang, Y., Mangat, R., Ruth, M., Shi, D., Jaeschke, A., and Russell, J. (2011). Chronic dietary n-3 PUFA intervention improves dyslipidaemia and subsequent cardiovascular complications in the JCR:LA-cp rat model of the metabolic syndrome. *Br. J. Nutr.* 105, 1572–1582.
- Marchioli, R., Barzi, F., Bomba, E., Chieffo, C., Di Gregorio, D., Di Mascio, R., Franzosi, M. G., Geraci, E., and Levantesi, G. (2002). Early protection against sudden death by n-3 polyunsaturated fatty acids after myocardial infarction: Time-course analysis of the results of the Gruppo Italiano per lo Studio della Sopravvivenza nell'Infarto Miocardico (GISSI)-Prevenzione. *Circulation* **105**, 1897–1903.
- Mozaffarian, D., Shi, P., Morris, J. S., Spiegelman, D., Grandjean, P., Siscovick, D. S., Willett, W. C., and Rimm, E. B. (2011). Mercury exposure and risk of cardiovascular disease in two U.S. cohorts. *N. Engl. J. Med.* **364**, 1116–1125.
- Murphy, R., Mourtzakis, M., Chu, Q., Baracos, V., Reiman, T., and Mazurak, V. (2011). Nutritional intervention with fish oil provides a benefit over standard of care for weight and skeletal muscle mass in patients with nonsmall cell lung cancer receiving chemotherapy. Cancer 117, 1775–1782.
- Neschen, S., Moore, I., Regittnig, W., Yu, C. L., Wang, Y., Pypaert, M., Petersen, K. F., and Shulman, G. I. (2002). Contrasting effects of fish oil and safflower oil on hepatic peroxisomal and tissue lipid content. Am. J. Physiol. Endocrinol. Metab. 282, E395–E401.
- Niu, J. L. and Kolattukudy, P. E. (2009). Role of MCP-1 in cardiovascular disease: Molecular mechanisms and clinical implications. *Clin. Sci.* **117**, 95–109.
- Oh, D., Talukdar, S., Bae, E., Imamura, T., Morinaga, H., Fan, W., Li, P., Lu, W., Watkins, S., and Olefsky, J. (2010). GPR120 is an omega-3 fatty acid receptor mediating potent anti-inflammatory and insulin-sensitizing effects. *Cell* **142**, 687–698.
- Oster, R. T., Tishinsky, J. M., Yuan, Z., and Robinson, L. E. (2010). Docosahexaenoic acid increases cellular adiponectin mRNA and secreted adiponectin protein, as well as PPARγ mRNA, in 3T3-L1 adipocytes. *Appl. Physiol. Nutr. Metab.* **35**, 783–789.
- Ott, J., Hiesgen, C., and Mayer, K. (2011). Lipids in critical care medicine. *Prostaglandins Leukot*. Essent. Fatty Acids 85, 267–273.
- Raitt, M. H., Connor, W. E., Morris, C., Kron, J., Halperin, B., Chugh, S. S., McClelland, J., Cook, J., and MacMurdy, K. (2005). Fish oil supplementation and risk of ventricular tachycardia and ventricular fibrillation in patients with implantable defibrillators: A randomized controlled trial. *JAMA* 293, 2884–2891.
- Rondanelli, M., Giacosa, A., Opizzi, A., Pelucchi, C., La Vecchia, C., Montorfano, G., Negroni, M., Berra, B., Politi, P., and Rizzo, A. (2011). Long chain omega 3 polyunsaturated fatty acids supplementation in the treatment of elderly depression: Effects on depressive symptoms, on phospholipids fatty acids profile and on health-related quality of life. *J. Nutr. Health Aging* 15, 37–44.
- Ruan, H. and Lodish, H. F. (2003). Insulin resistance in adipose tissue: Direct and indirect effects of tumor necrosis factor-alpha. *Cytokine Growth Factor Rev.* **14**, 447–455.
- Sampath, H. and Ntambi, J. M. (2004). Polyunsaturated fatty acid regulation of gene expression. Nutr. Rev. 62, 333–339.
- Schwab, J. M., Chiang, N., Arita, M., and Serhan, C. N. (2007). Resolvin E1 and protectin D1 activate inflammation-resolution programmes. *Nature* **447**, 869–874.

- Sessler, A. M., Kaur, N., Palta, J. P., and Ntambi, J. M. (1996). Regulation of stearoyl-CoA desaturase 1 mRNA stability by polyunsaturated fatty acids in 3T3-L1 adipocytes. *J. Biol. Chem.* **271**, 29854–29858.
- Simopoulos, A. P. (2002). The importance of the ratio of omega-6/omega-3 essential fatty acids. *Biomed. Pharmacother.* **56**, 365–379.
- Suzuki, M., Noda, K., Kubota, S., Hirasawa, M., Ozawa, Y., Tsubota, K., Mizuki, N., and Ishida, S. (2010). Eicosapentaenoic acid suppresses ocular inflammation in endotoxininduced uveitis. *Mol. Vis.* 16, 1382–1388.
- Thies, F., Garry, J. M., Yaqoob, P., Rerkasem, K., Williams, J., Shearman, C. P., Gallagher, P. J., Calder, P. C., and Grimble, R. F. (2003). Association of n-3 polyunsaturated fatty acids with stability of atherosclerotic plaques: A randomised controlled trial. *Lancet* 361, 477–485.
- Virtanen, J. K., Voutilainen, S., Rissanen, T. H., Mursu, J., Tuomainen, T. P., Korhonen, M. J., Valkonen, V. P., Seppänen, K., Laukkanen, J. A., and Salonen, J. T. (2005). Mercury, fish oils, and risk of acute coronary events and cardiovascular disease, coronary heart disease, and all-cause mortality in men in eastern Finland. *Arterioscler. Thromb. Vasc. Biol.* 25, 228–233.
- Wall, R., Ross, R., Fitzgerald, G., and Stanton, C. (2010). Fatty acids from fish: The anti-inflammatory potential of long-chain omega-3 fatty acids. *Nutr. Res.* **68**, 280–289.
- Wan, J. B., Huang, L. L., Rong, R., Tan, R., Wang, J., and Kang, J. X. (2010). Endogenously decreasing tissue n-6/n-3 fatty acid ratio reduces atherosclerotic lesions in apolipoprotein E-deficient mice by inhibiting systemic and vascular inflammation. *Arterioscler. Thromb. Vasc. Biol.* **30**, 2487–2494.
- Whelan, J. and Rust, C. (2006). Innovative dietary sources of n-3 fatty acids. *Annu. Rev. Nutr.* **26**, 75–103.
- White, P. J., Arita, M., Taguchi, R., Kang, J. X., and Marette, A. (2010). Transgenic restoration of long-chain n-3 fatty acids in insulin target tissues improves resolution capacity and alleviates obesity-linked inflammation and insulin resistance in high-fat-fed mice. *Diabetes* 59, 3066–3073.
- Wortman, P., Miyazaki, Y., Kalupahana, N. S., Kim, S., Hansen-Petrik, M., Saxton, A. M., Claycombe, K. J., Voy, B. H., Whelan, J., and Moustaid-Moussa, N. (2009). n3 and n6 polyunsaturated fatty acids differentially modulate prostaglandin E secretion but not markers of lipogenesis in adipocytes. *Nutr. Metab. (Lond)* 6, 5.
- Wu, M., Harvey, K., Ruzmeto, N., Welch, Z., Sech, L., Jackson, K., Stillwell, W., Zaloga, G., and Siddiqui, R. (2005). Omega-3 polyunsaturated fatty acids attenuate breast cancer growth through activation of a neutral sphingomyelinase-mediated pathway. *Int. J. Cancer* 117, 340–348.
- Younce, C. W., Azfer, A., and Kolattukudy, P. E. (2009). MCP-1 (monocyte chemotactic protein-1)-induced protein, a recently identified zinc finger protein, induces adipogenesis in 3T3-L1 pre-adipocytes without peroxisome proliferator-activated receptor gamma. J. Biol. Chem. 284, 27620–27628.
- Zhao, Y., Joshi-Barve, S., Barve, S., and Chen, L. (2004). Eicosapentaenoic acid prevents LPS-induced TNF-alpha expression by preventing NF-kappa B activation. *J. Am. Coll. Nutr.* **23**, 71–78.
- Zhu, J., Yong, W., Wu, X., Yu, Y., Lv, J., Liu, C., Mao, X., Zhu, Y., Xu, K., and Han, X. (2008). Anti-inflammatory effect of resveratrol on TNF-alpha-induced MCP-1 expression in adipocytes. *Biochem. Biophys. Res. Commun.* 369, 471–477.